

# Chapter 6

## Indole-3-Carbinol and Its Role in Chronic Diseases

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**Abstract** Indole-3-carbinol (I3C), a common phytochemical in cruciferous vegetables, and its condensation product, 3,3'-diindolylmethane (DIM) exert several biological activities on cellular and molecular levels, which contribute to their well-recognized chemoprevention potential. Initially, these compounds were classified as blocking agents that increase drug-metabolizing enzyme activity. Now it is widely accepted that I3C and DIM affect multiple signaling pathways and target molecules controlling cell division, apoptosis, or angiogenesis deregulated in cancer cells. Although most of the current data support the role of I3C and DIM in prevention of hormone-dependent cancers, it seems that their application in prevention of the other cancer as well as cardiovascular disease, obesity, and diabetes reduction is also possible. This chapter summarizes the current experimental data on the I3C and DIM activity and the results of clinical studies indicating their role in prevention of chronic diseases.

**Keywords** Indole-3-carbinol · DIM · Signaling pathways · Chronic diseases · Animal models · Dietary intervention trials

### 6.1 Introduction

The plant family *Cruciferae*, particularly members of the genus *Brassica*, like cabbage, broccoli, cauliflower, Brussels sprouts, kale, bok choy are rich sources of sulfur-containing glucosinolates. These secondary products of plant metabolism include, among others, glucobrassicin and neoglucobrassicin. When plant tissue is disrupted, an endogenous thioglucosidase (myrosinase) is activated and converts

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glucobrassicin and other indolylic glucosinolates to indoles, principally to indole-3-carbinol (I3C) [120].

Since the first reports on possible anti-carcinogenic activity of I3C [122] numerous preclinical studies have confirmed the chemopreventive properties of this compound by preventing, inhibiting, and reversing the progression of cancer. Moreover, preliminary clinical trials have shown that I3C is a promising agent protecting against hormone-dependent as well as hormone-independent human cancers [47].

Thus, it is not surprising that there are many marked diet supplements containing I3C. Several mechanistic studies have been performed in order to elucidate the mechanism of pleiotropic activity of I3C.

This chapter summarizes the current knowledge on the possible interference of I3C with signaling pathways *in vitro* and in animal models, as well as its application in prevention of chronic diseases.

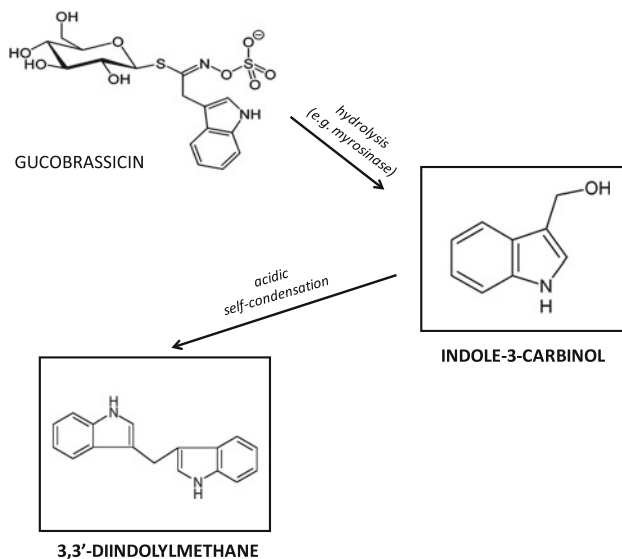
## 6.2 Physicochemical Properties and Pharmacokinetics of Indole-3-Carbinol

Among the indoles, generated upon ingestion of cruciferous vegetables, only I3C (IUPAC: 1H-indol-3-ylmethanol) is commercially available as an off-white solid. Basic physical and chemical properties of I3C are summarized in Table 6.1.

I3C is chemically unstable in acidic conditions, *in vitro* in cell cultures and *in vivo* in the stomach environment. In such conditions, I3C may rapidly condense into a series of oligomeric products, of which a dimer, 3,3,-diindolylmethane (DIM), is considered the most bioactive product (Fig. 6.1) [2, 115]. Several pharmacokinetics studies, performed mostly in animal models, have been conducted for I3C and its condensation products [5, 6, 34, 39, 105]. When rainbow trout has been administered with radiolabeled [5-<sup>3</sup>H]-indole-3-carbinol, 40 % of total radioactivity was found in the liver extracts as DIM [34]. Upon oral administration of 250 mg/kg to mice, the I3C was rapidly absorbed and distributed into variety of tissues and body fluids (e.g., liver, kidney, lung, heart, brain, and plasma) with highest concentrations in liver and kidney, but with rapid clearance (concentrations below the limit of detection within 1 hour after administration). In the same experiment, DIM was detected in plasma at 15 min and was still quantifiable after 6 h with a peak at 2 h after I3C dosing [5, 6]. DIM was also found in stomach tissue and contents,

**Table 6.1** Physical and chemical properties of I3C (ALOGPS, [www.pubchem.com](http://www.pubchem.com); accessed Dec 26, 2015)

Stability	Off-white powder
Molecular weight	147.17386 g/mol
Melting range	96–99 °C
Storage temperature	2–8 °C
Stability	2–80 °C, considered stable
Water solubility	3.75 mg/ml, mixes with water

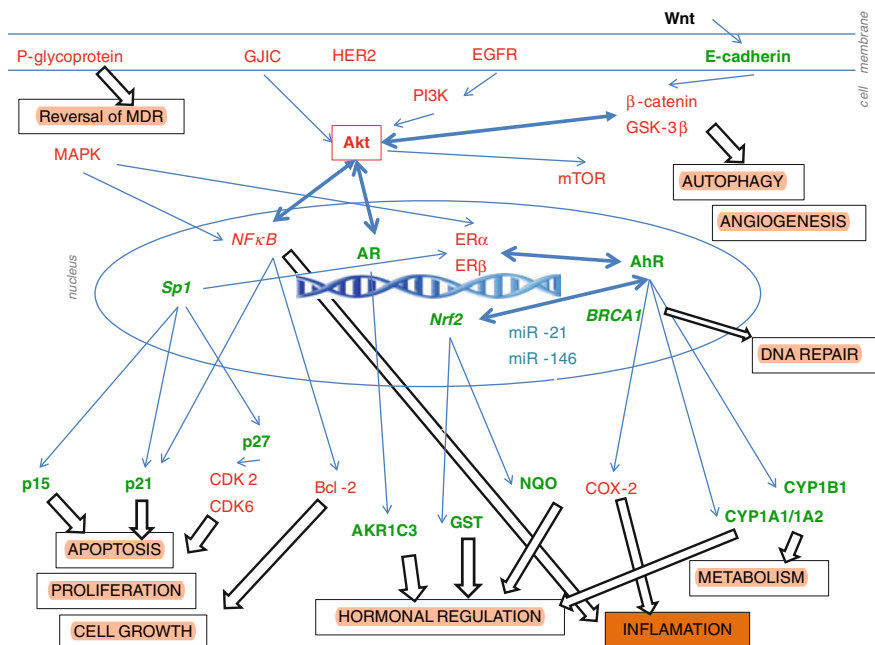


**Fig. 6.1** Molecular structure and formation of I3C and DIM

intestines, and liver after 1 h following oral administration of I3C to rats [39]. In human volunteers intervention trial DIM was detected in plasma within 8 h following of 400 mg I3C oral dose [9]. In a phase I clinical trial in women, no I3C was found in the plasma after administration of a single dose of up to 1200 mg or multiple-doses at 400 mg provided twice daily for 4 weeks, and DIM was the only detectable I3C-derived compound in plasma [105]. Fujioka et al. [47] have found that urinary DIM level after uptake of I3C from Brussels sprouts or cabbage is a biomarker of glucobrassicin exposure in humans. All these results support the suggestion that I3C serves as the prodrug rather than the therapeutic agent itself. In this regard, purified I3C as treatment agent used in in vitro models seems to be somewhat contradictory, because there is no certainty that any metabolism of DIM in cells occur. Thus, in this chapter the biological activity and the role in chronic diseases will refer not only to I3C but also to DIM, its major condensation product in humans.

### 6.3 Modulation of Cell Signaling Pathways by Indole-3-Carbinol

I3C affects multiple signaling pathways and target molecules controlling cell division, apoptosis or angiogenesis deregulated in cancer cells. Figure 6.2 presents the overview of the signaling pathways and possible crosstalks influenced by I3C or DIM. One of the major pathways targeted by I3C is phosphoinositide 3-kinase



**Fig. 6.2** Signaling pathways and proposed crosstalks (↔) affected by I3C/DIM; red-inhibition, green-induction (AhR-aryl hydrocarbon receptor, AKR-aldol-keto reductase, Akt- protein kinase B, AR-androgen receptor, BRCA-breast cancer tumor suppressor gene, CDK-cyclin-dependent kinase, COX-cyclooxygenase, EGFR-epidermal growth factor receptor, ER-estrogen receptor, GJIC-Gap junctional intracellular communication, GSK-glycogen synthase kinase, GST-glutathione-S-transferases, HER-receptor tyrosine-protein kinase erbB-2, MAPK-mitogen-activated protein kinases, MDR-multidrug resistance, NFκB-nuclear factor kappa-light-chain-enhancer of activated B cells, Nrf2-nuclear transcription factor 2, NQO-NAD (P)H:quinone oxidoreductases, PI3K-phosphoinositide 3-kinase, Sp1-Sp1 transcription factor)

(PI3K)—protein kinase B (Akt)—mammalian target of rapamycin (mTOR) signaling pathway. This pathway is a cascade of events that plays a key role in the broad variety of physiological and pathological processes. PI3K/Akt/mTOR signaling pathway is one of the most frequently affected target in all sporadic human cancers, and it has been estimated that mutations in the individual component of this pathway account for as much as 30 % of all known human cancers [83, 112].

Akt is a serine/threonine protein kinase functioning downstream of PI3K in response to mitogen or growth factor stimulation. The inhibition of phosphorylation and subsequent activation of Akt kinase by I3C or DIM was described in prostate and breast cancer cells. In addition, I3C abrogated epidermal growth factor (EGF)-induced activation of Akt in prostate cancer cells. Furthermore, the known downstream modulators of the Akt/PI3K cell survival pathway, Bcl-x(L), and BAD proteins showed decreased expression after I3C treatment [24, 49]. Several genes that mediate processes involved in carcinogenesis are regulated by transcription

factor NF- $\kappa$ B. It plays a central role in general inflammation as well as immune responses, although recent evidences suggest that its role in these processes is more complex [73]. The enhanced activation and expression of NF- $\kappa$ B is linked to development and progression of human cancers as well as in the acquisition of drug-resistant phenotype in highly aggressive malignancies [3, 109].

Activation through Akt is important for many tumorigenic functions of NF- $\kappa$ B. Several studies showed that both indoles inhibited PI3K/Akt/mTOR/NF- $\kappa$ B signaling (reviewed in [4]).

It was also known that there is a crosstalk between Akt and Wnt signaling pathways through the signal communication between glycogen synthase kinase 3 (GSK-3 $\beta$ ) and  $\beta$ -catenin, two of the important molecules in Akt and Wnt pathways, respectively [109]. It was found that DIM significantly increased the phosphorylation of  $\beta$ -catenin, and inhibited  $\beta$ -catenin nuclear translocation [77] suggesting that DIM could also downregulate the activation of Wnt signaling.

Mitogen-activated protein kinases (MAPK) are involved in cellular response to a diverse array of stimuli and regulate several cell functions including differentiation, mitosis, cell survival, and apoptosis [100]. Downregulation of the expression of MAP2K3, MAP2K4, MAP4K3, and MAPK3 by I3C and DIM was described suggesting their inhibitory effects on MAPK pathway [76]. It was also reported that the effects of DIM were mediated by crosstalk between protein kinase A and MAPK signaling pathways [75].

Gap junctional intracellular communication (GJIC) also involved in regulating cell proliferation, differentiation and apoptosis is modulated by gap junction channels via broad variety of endogenous and exogenous agents [90]. I3C was reported to prevent H<sub>2</sub>O<sub>2</sub>-induced inhibition of GJIC by the inactivation of Akt in rat liver epithelial cells [57].

Several studies have focused on the potential effects of I3C and DIM on the proliferation and induction of apoptosis in human prostate or breast cancer cell lines. Cell G<sub>1</sub> cycle arrest of breast cancer cells by I3C was related to inhibition of the expression of cyclin-dependent kinase-6 (CDK6) independently of estrogen receptor signaling [30]. Moreover, the inhibition of CDK6 expression in human MCF7 breast cancer cells was further explained by disrupting Sp1 transcription factor interaction with CDK6 gene promoter [32].

In prostate cancer cell lines, the induction of apoptosis by I3C was p53-independent [94]. The induction of p21<sup>WAF1</sup> expression by DIM was also independent of p53 status [55]. Pro-apoptotic effect of DIM in HER2/Neu over-expressing breast cancer cells was connected to inhibition of HER2/Neu activity [87]. We have also found a decreased expression of HER2/Neu in estrogen independent MDA-MB-231 breast cancer cells treated with either I3C or DIM (Licznarska et al. unpublished data).

I3C has been known to be a negative regulator of estrogens, while DIM a negative regulator of androgens. I3C inhibited the transcriptional activity of ER $\alpha$ , the estradiol-activated ER $\alpha$  signaling, and the expression of the estrogen-responsive genes [89]. Since I3C and DIM could also inhibit the proliferation of ER-negative breast cancer cells, it is suggested that antitumor activities of indoles could be ER

independent. DIM was found to be an antagonist of AR, inhibiting androgen-induced AR translocation to the nucleus [74]. DIM also enhanced expression of Aldo-keto reductase 1C3 (AKR1C3), an enzyme responsible for inactivation of 5 $\alpha$ -dihydrotestosterone and decreasing estrogen levels in mammary gland and eliminating active androgens from the prostate [92, 101].

Several lines of evidence indicate possible crosstalk between ER $\alpha$  and AhR signaling pathways [96]. It was shown that I3C triggers AhR-dependent ER $\alpha$  protein degradation in MCF7 breast cancer cell line disrupting an ER $\alpha$ -GATA3 transcription factor cross-regulatory loop. This led to ablation of ER $\alpha$  expression and loss of ER $\alpha$ -responsive proliferation [86].

Our studies showed that both indoles upregulated AhR and downregulated ER $\alpha$  expression in non-tumorigenic MCF10A and tumorigenic MCF7 breast epithelial cells [114]. Upregulation of AhR was also observed in ER-negative MDA-MB-231 cells. This observation is important since increased expression and activation of AhR result in induction of CYPs involved in estrogen metabolism [113].

Both I3C and DIM elicited an inhibition of cell adhesion, migration and invasion in the breast cell lines of different ER status [89]. This appeared to be due, in part, to I3C-induced upregulation of the tumor suppressor gene *BRCA1* and other proteins involved in DNA repair, like RAD51 [105].

Recent studies show that AhR and ER $\alpha$  interact with and modulate the activity of Nrf2 [52]. This transcription factor plays an essential role in cellular protection against electrophiles and oxidative stress by upregulating expression of phase II detoxifying enzymes, including glutathione-S-transferases (GST) and NAD(P)H:quinone oxidoreductases (NQO) [81]. Significant increase of the Nrf2 transcript level in MCF7 and MDA-MB-231 breast cancer cell lines was observed after treatment with I3C. Moreover, increase of Nrf2 expression was correlated with enhanced expression of NQO1 or GSTP in MCF7 cells and MDA-MB-231 cells respectively [114].

Recently, a plethora of evidence has demonstrated that epigenetic alterations, such as DNA methylation, histone modifications, and non-coding miRNAs, consistently contribute to carcinogenesis, and dietary phytochemicals, including glucosinolate derivatives, have the potential to alter a number of these epigenetic events [46, 119].

In this regard, DIM was reported to decrease promoter methylation of Nrf2 in vitro in TRAMP C1 mouse prostate cell line and in vivo in TRAMP mice prostate tumors. This effect was at least in part related to decreased expression and activity of DNA methyltransferases [130]. Moreover, genome-wide promoter methylation in normal and cancer prostate cells showed broad and complex effects on DNA methylation profiles reversing many of the cancer associated methylation alterations, including aberrantly methylated genes that are dysregulated or are highly involved in cancer progression [129].

Histone modifications by DIM were found in several studies. Decreased histone deacetylases (HDACs) protein expression were observed in human colon, breast,

and prostate cancer cell lines, human colon cancer xenografts in nude mice and in mouse prostate cells, influencing apoptotic proteins, like p21, p27, and involved in inflammation COX-2 [46].

I3C and DIM were found to downregulate miR-21 and miRNA-146, respectively, in Panc-1 pancreatic cancer cells, which was related to induction of chemosensitivity to gemcitabine in these cells [78, 98]. In another study, I3C reversed the upregulation of miR-21 caused by lung carcinogen—vinyl carbamate in mice [88]. DIM upregulated miR-21 in breast cancer MCF7 cells resulting in reduced proliferation [62], and the let-7 family miRNA leading to inhibition of self-renewal and clonogenic capacity of prostate cancer cells [69, 70].

To sum up, both I3C and its dimer demonstrate pleiotropic effects on cell signaling and subsequently gene expression regulation. Some of these activities are summarized in Table 6.2.

**Table 6.2** Summary of the major biological effects of indole-3-carbinol (I3C) and its condensation product—3,3'-diindolylmethane (DIM)

Effect <sup>a</sup>	Target molecules	I3C		DIM	
		In vitro	In vivo	In vitro	In vivo
Induction of phase I enzymes	CYP1A1, CYP1A2, CYP1B1	+	+	+	
Induction of phase II enzymes	GST, NQO	+	+	+	+
Inhibition of DNA adducts formation	CYP1B1		+		
Anti-estrogenic activity	ER, AhR	+	+	+	
Anti-androgenic activity	AR	+		+	
Cell cycle arrest	p21, p27, CDK6	+		+	
Pro-apoptotic	Akt, NFκB, GSK3β, JNK	+		+	
Anti-angiogenic activity	E-cadherin, α-, β-, and γ-catenins, MMP9	+		+	+
Anti-proliferative activity	ERβ/ERα	+	+	+	
DNA repair	BRCA1, RAD51	+	+	+	
Reversal of MDR	P-glycoprotein	+	+		
Epigenetic modifications	DNMTs, HDACs, miRNAs			+	+
Anti-inflammatory activity	NFκB, COX-2	+	+	+	

AhR aryl hydrocarbon receptor, Akt protein kinase B, AR androgen receptor, BRCA breast cancer tumor suppressor gene, CDK cyclin-dependent kinase, DNMT DNA methyltransferase, ER-estrogen receptor, GSK glycogen synthase kinase, GST glutathione-S-transferases, HDAC histone deacetylases, JNK c-Jun N-terminal kinase, MDR multidrug resistance, MMP matrix metalloproteinase, NFκB nuclear factor kappa-light-chain-enhancer of activated B cells, NQO NAD(P)H:quinone oxidoreductases

<sup>a</sup>References in the text

## 6.4 Role of Indole-3-Carbinol in Chronic Diseases

The term “chronic diseases” appear under different names in different contexts. According to WHO this term suggests the following shared features:

- The chronic disease epidemics take decades to become fully established—they have their origins at young age;
- Given their long duration, there are many opportunities for prevention;
- They require a long-term and systematic approach to treatment (WHO Report 2015).

In this context, cardiovascular diseases, chronic respiratory diseases, diabetes along with cancer are mentioned. However, chronic character of cancer is not so obvious as in the case of the other illnesses. The idea of considering cancer as chronic disease emerged recently when it was noted that many cancers, while still very serious could be manageable chronic diseases with ongoing surveillance and/or treatment. While this vision has not yet become a reality for most forms of cancer, the past 10–20 years have brought about a marked acceleration in advance toward this goal. In this regard some types of metastatic breast cancer have become manageable over the long term, perhaps most famously with tamoxifen, which can slow or stop malignant cell growth in many women with estrogen-dependent cancer by blocking hormone receptors on tumor cells. Moreover, a new class of aromatase (*CYP19*) inhibitors that target estrogen production was developed, which seem to provide better results than tamoxifen [127]. I3C can also inhibit the expression of aromatase as well as CYP isoforms involved in estrogen metabolism. Therefore, I3C, as well as its condensation product DIM, appear to be a promising agent for the prevention or recurrence of human tumors, particularly hormone-dependent cancers. There are also data suggesting that I3C or DIM might also be useful for prevention or recurrence of cardiovascular diseases, diabetes, or recurrent respiratory papillomatosis, as well as obesity.

### 6.4.1 *The Role of I3C in Cancer Controlling*

It is almost 40 years after Lee Wattenberg and William Loub [122] for the first time reported that I3C inhibited chemically induced breast and forestomach neoplasia in rodents. Since then the antitumor activity of dietary I3C has been widely studied and the inhibition of the development of other cancer types, including liver [95], lung [63], and prostate [110] have been demonstrated.

Breast cancer is the most common and the leading cause of cancer mortality among women worldwide. The preventive efficacy of I3C on mammary carcinoma first observed in animal models, was confirmed in many mechanistic studies in cell cultures and was supported by epidemiological studies with cruciferous vegetables and their extracts or juices [117, 120].



One of the most important risk factor of breast tumors are estrogens, which are classified as carcinogenic in humans [58]. These steroid hormones may contribute to breast cancer development in two ways: (i) acting as promoters by stimulating cell proliferation, (ii) inducing genotoxicity through the reaction of their active metabolites with DNA thus acting as tumor initiators.

Experimental studies *in vitro* in breast epithelial cell lines showed that I3C, DIM, and cabbage juices induce CYP450 genes *CYP1A1*, *1A2*, *1B1* encoding the key enzymes of estrogen catabolism. The profile of metabolites was in favor to 2-hydroxyestrogens being noncarcinogenic in comparison to estradiol and 4-hydroxy derivatives [80, 114]. This anti-estrogenic activity of I3C could be explained by the induction of AhR receptor showed in other studies [114].

Moreover, I3C, DIM, and cabbage juices are capable of upregulating phase II detoxifying enzymes, including GST and NQO in breast cancer cell lines [114, 120]. Upregulation of GSTs and NQO1 by I3C was correlated with increased levels of Nrf2, in benign MCF7 and aggressive MDA-MB-231 breast cancer cell lines. Thus, it may be assumed that I3C protects against estrogen-associated carcinogenesis by removal of the genotoxic metabolites of estrogens.

Simultaneously, I3C and DIM influenced *in situ* production of estrogens in breast epithelial MCF7 cancer cells by reducing the expression of aromatase (*CYP19*), the enzyme that synthesizes estrogens by converting C19 androgens into aromatic C18 estrogenic steroids [80]. Several studies have shown that there is an overexpression of aromatase gene in breast cancer tissue [82]. Interestingly, the potential of I3C to reduce estrogenic activity in the breast cancer cells was confirmed by other mechanisms, particularly via decreasing the AKR1C3 expression mentioned in the previous section. In the mammary gland where enzyme converts androstenedione to testosterone—one of the aromatase substrates [92, 101].

Besides the interference with estrogens metabolism pathways, I3C also affects DNA repair, the cell cycle progression and apoptosis in breast cancer cell lines. In this regard it was shown that I3C induces BRCA1 expression and that both I3C and BRCA1 inhibited estrogen (E2)-stimulated ER $\alpha$  activity in human breast cancer cells [44]. BRCA1 is DNA repair factor involved in repair of DNA double-strand breaks. *BRCA1* gene expression is reduced or completely silenced in a significant proportion of sporadic breast cancer because of hypermethylation of the gene promoter [106, 126]. Thus, I3C-induced BRCA1 expression and inhibition of estrogen-stimulated ER $\alpha$  activity by I3C and BRCA1 showed by some studies could be one of the antitumor activity of the indole [44].

Several lines of evidence suggest I3C ability to arrest cell cycle in breast cancer cells. In this regard I3C was reported to inhibit CDK2 activity in breast cancer MCF7 cell line [48]. Moreover, both I3C and DIM upregulated CDK inhibitors p21 and p27, although in a very high concentration of 200  $\mu$ M. Activity of this protein is especially critical during the G<sub>1</sub> to S phase transition. Consequently, the ability to arrest G<sub>1</sub> phase by I3C was shown [30]. Other studies suggest the p53 phosphorylation by I3C leading to release p53 and inducing the p21 CDK inhibition and G<sub>1</sub> cell cycle arrest [18, 86]. Importantly, treatment with I3C and tamoxifen ablated expression of the phosphorylated retinoblastoma protein (Rb), an endogenous

substrate for the G<sub>1</sub> CDKs, whereas either agent alone only partially inhibited endogenous Rb phosphorylation [31]. Several studies showed that I3C and its derivatives are potent inducers of apoptosis in both ER-positive and ER-negative breast cancer cells [30–32, 50, 80, 103]. Estrogens, particularly estradiol have been also implicated as a cofactors in human papillomavirus (HPV)-mediated cervical cancer, both in animal models and in women using oral contraceptives [72]. Interestingly, it was found that estradiol protects cervical cancer cells treated with DNA-damaging agents such as UVB, mitomycin-C, and cisplatin, from apoptotic death. I3C was able to overcome the anti-apoptotic effect of estradiol but only in higher concentrations. Treatment with I3C resulted in loss of the survival protein Bcl-2. However, the amount of apoptosis versus survival and the level of Bcl-2 depended on the I3C/estradiol ratio [22]. In HPV16-transgenic mice, which develop cervical cancer after chronic estradiol exposure, apoptotic cells were detected in cervical epithelium only in mice exposed to estradiol and fed on I3C [20].

Experiments in which cervical cancer HeLa and SiHa cell lines were used, demonstrated that DIM also exerts antitumor effects on these cells through its anti-proliferative and pro-apoptotic roles, especially for SiHa cells. The molecular mechanism for these effects may be related to its regulatory effects on MAPK and PI3K pathway and apoptosis proteins. Thus, DIM may be considered a preventive and therapeutic agent against cervical cancer [135]. The ability of inhibiting spontaneous occurrence of endometrial adenocarcinoma and preneoplastic lesions by I3C was also demonstrated in female Donryu rats. It was suggested that this effect was due to induction by I3C estradiol 2-hydroxylation [68]. On the other hand, promotion of endometrial adenocarcinoma in same strains of rats initiated with N-ethyl-N'-nitro-N-nitrosoguanidine by I3C was described [133]. DIM was also found to have a potent cytostatic effect in cultured human Ishikawa endometrial cancer cells. This effect was related to the stimulation of TGF- $\alpha$  expression and activation of TGF- $\alpha$  signal transduction pathway [75].

Another hormone-dependent cancer, which might be affected by I3C, is prostate cancer, one the most prevalent malignancy in men worldwide and the second leading cause of male death in Western countries [16]. Androgens play a critical role in prostate cancer cells growth and survival. Androgens bind to the androgen receptor (AR), a steroid nuclear receptor, which is translocated into the nucleus and binds to AREs in the promoter regions of target genes to induce cell proliferation and apoptosis. Approximately 80–90 % of prostate cancers are dependent on androgen at initial diagnosis, and endocrine therapy of prostate cancer is directed toward the reduction of plasma androgens and inhibition of AR [37, 54]. It was demonstrated that both I3C and DIM are able to downregulate AR signaling [74], but only DIM was shown to be a strong antagonist of AR and inhibitor of its translocation to the nucleus [30, 74].

Similarly as in the case of breast cancer cells, I3C and its derivatives also affect cell cycle progression and induce apoptosis. In this regard, cell cycle arrest at G<sub>1</sub> checkpoint in different human prostate carcinoma cell lines by I3C and DIM was described [2]. In LNCaP prostate cancer cells I3C selectively inhibited the expression of CDK6 protein and transcripts and stimulated the production of the

p16 CDK inhibitor. In vitro protein kinase assays revealed inhibition by I3C CDK2 enzymatic activity and the relatively minor downregulation of CDK4 enzymatic activity [134].

In PC-3 cell line induction of G<sub>1</sub> cell cycle arrest by I3C due to the upregulation of p21(WAF1) and p27(Kip1) CDK inhibitors, followed by their association with cyclin D1 and E and downregulation of CDK6 protein kinase levels and activity was suggested. In addition, I3C inhibited the hyperphosphorylation of the retinoblastoma (Rb) protein in PC-3 cells. Induction of apoptosis was also observed in this cell line when treated with I3C. Thus, it was suggested that I3C inhibits the growth of PC-3 prostate cancer cells by inducing G<sub>1</sub> cell cycle arrest leading to apoptosis, and regulates the expression of apoptosis-related genes [23]. Further studies showed that I3C-induced apoptosis is partly mediated by the inhibition of Akt activation, resulting in the alterations in the downstream regulatory molecules of Akt activation in PC-3 cells [24]. In the case of DIM an inhibition of a crosstalk between Akt and NF- $\kappa$ B [12], leading to cell cycle arrest and induction of apoptosis was also described. DIM significantly decreased cellular histone deacetylase HDAC2 protein level in androgen sensitive LNCaP and androgen insensitive PC-3 cell lines [10]. In all these studies a formulated DIM (BR-DIM) with higher bioavailability was used and was able to induce apoptosis and inhibit cell growth, angiogenesis, and invasion of prostate cancer cells [21].

The potential protective activity of I3C and DIM against prostate cancer was confirmed by microarray analysis, which showed the modulation of the expression of many genes related to the control of carcinogenesis and cell survival as effect of indoles treatment of PC-3 cells [76]. It was also demonstrated by several groups that I3C and DIM may improve the therapeutic effect of conventional chemotherapy of prostate cancer [44, 71].

Besides the hormone-dependent cancers, both indoles can affect the development of some other cancers. In this regard, it was shown that I3C and DIM induced apoptosis in colorectal cancer cell lines [13, 67, 84]. Interestingly, an effective inhibition of Akt and inactivation of mTOR was observed as a result of combined treatment with I3C and genistein in HT29 colon cancer cells, leading to induction of apoptosis and autophagy [93].

Anti-carcinogenic activity of I3C was demonstrated in carcinogen-induced lung cancer in mice [63, 64]. Anti-proliferative effects of I3C and DIM in human bronchial epithelial cells (HBEC) and A549 adenocarcinomic human alveolar basal epithelial cells related to marked reductions in the activation of Akt, extracellular signal-regulated kinase and NF- $\kappa$ B were also described [65].

Moreover, upregulation of several miRNAs induced by chemical carcinogen was reversed by I3C in mice and rats lung tumors [46, 59].

The signal transducer and activator of transcription 3 (STAT3) is a latent transcription factor required in proliferation and differentiation. The constitutive activation of STAT3 in human pancreatic carcinoma specimens but not in normal tissues was shown. Activation of STAT3 was also found in pancreatic tumor cell lines and was inhibited by I3C although in relatively high concentration (10  $\mu$ M) along with induction of apoptosis [79]. Apoptosis in pancreatic cancer cells was

also induced by DIM as a result of endoplasmic reticulum stress-dependent upregulation of death receptor 5 [1]. More recent studies showed downregulation of miRNA-21 and miRNA-221 as a result of I3C or DIM treatment of pancreatic cancer cells. As upregulation of these miRNAs is characteristic for more aggressive pancreas cancer, it was suggested that combination of I3C or DIM with conventional chemotherapeutics may increase the chemosensitivity to certain drugs in resistant pancreatic cancer cells [98, 111].

There are also reports showing the anti-proliferative effect, G1 cell cycle and induction of apoptosis in thyroid cancer cells by I3C and DIM [116]. However, earlier reports indicated the enhancement of thyroid gland neoplastic development by I3C in rat medium-term multi-organ carcinogenesis model [66]. UVB-induced mouse skin tumors were reduced in mice fed on I3C [29].

Importantly it was also shown that I3C may overcome multiple drug resistance by downregulation of MDR1-expression in murine melanoma cells and leukemia cells [7, 8, 28].

These observations further support the possible application of I3C or DIM as potential adjuvant therapeutics in conventional chemotherapy of several cancers.

Finally, it must be pointed out that although I3C was shown to have anti-carcinogenic activity in various animal models, at the same time animals studies have also shown a tumor-promoting activity, when animals were exposed to I3C after exposure to carcinogens [131, 133]. This aspect has to be clarified in long-term studies.

#### 6.4.2 **Cardiovascular Diseases**

Cardiovascular diseases still remain the primary cause of death worldwide. One of the proposed approaches to reduce the high global incidence is the consumption of vegetables and fruits containing biologically active components or phytochemical supplements. Although the most attention was paid to resveratrol, components of cruciferous vegetables, particularly *Brassica oleracea*, were also considered a potential dietary phytochemicals reducing risk of CVDs [97].

In this regard hypocholesterolemic properties of I3C were reported in mice provided with cholesterol-supplemented diet to which I3C were added. Since *in vitro* experiments revealed that I3C and its condensation products effectively inhibited the enzyme acyl-CoA:cholesterol acyltransferase (ACAT), which is responsible for the conversion of free cholesterol to the cholesteryl ester, the hypocholesterolemic effect of I3C in mice was likely mediated by the inhibition of ACAT [42]. Such mechanism was further confirmed in HepG2 cells. As a result of treatment with I3C the decreased cholesteryl ester synthesis was associated with significantly decreased ACAT gene expression and activity [85].

Moreover, antiplatelet and antithrombotic activity of I3C was shown in *in vitro* and *in vivo* studies. I3C significantly inhibited collagen-induced platelet aggregation in human platelet-rich plasma and suppressed the death of mice with

pulmonary thrombosis induced by intravenous injection of collagen and epinephrine [99].

The protective activity of I3C in heart failure and vascular proliferative disease was also reported. In this regard, it was shown that I3C can suppress the proliferation of cultured vascular smooth muscle cells and neointima formation in a carotid injury model via the Akt/GSK3 $\beta$  pathway [51]. Vascular smooth muscle cells are the principal cell types involved in the pathogenesis of atherosclerosis and restenosis after percutaneous coronary intervention [43]. Thus, it was suggested that I3C may be a part of new therapeutic strategy for vascular proliferative diseases as well as heart failure. The latter suggestion was supported by the results of the studies using aortic banding (AB) mouse model, which showed that I3C prevented and reversed cardiac remodeling induced by AB. This effect was mediated by AMPK- $\alpha$  and extracellular signal-regulated kinases 1/2 (ERK1/2) [36]. Since AMPK acts as important energy sensor, attenuation of cardiac remodeling in mice was associated with improved myocardial energy metabolism [36].

### 6.4.3 *Obesity and Diabetes*

Chronic inflammatory disease initiated in adipose tissue might lead to obesity-related insulin resistance and may contribute to an increased risk of diabetes [132]. It might be assumed that anti-inflammatory phytochemicals may protect against both diseases. Thus, I3C was also proposed as a potential preventive agent against obesity and metabolic disorders. In this regard, I3C treatment in diet-induced obesity (DIO) mice model decreased body weight and fat accumulation and infiltrated macrophages in epididymal adipose tissue. These effects were associated with improved glucose tolerance and with modulated expression of adipokines and lipogenic-associated gene products, including acetyl coenzyme A carboxylase and peroxisome proliferator-activated receptor- $\gamma$  (PPAR $\gamma$ ) [19]. The reduced level of inflammatory biomarkers was also confirmed in co-culture of adipocytes with macrophages treated with I3C [19].

I3C was also capable to normalize tissue expression of genes related to thermogenesis upregulated by high-fat diet, namely uncoupling proteins 1 and 3, PPAR $\alpha$ , PPAR $\gamma$  coactivator 1 $\alpha$  [26]. The observed improvement of adipogenesis by I3C could be due to activation of sirtuine SIRT1 [27]. These findings suggest that I3C has a potential benefit in preventing obesity and metabolic disorders, and the action for I3C in vivo may involve multiple mechanisms including decreased adipogenesis and inflammation, along with activated thermogenesis.

Little is known about the possible modulation of different types of diabetes by I3C. Nevertheless, in recent studies with the genetically modified mice (C57BL/6J mice) that closely simulated the metabolic abnormalities of the human disease after the administration of high-fat diet, both I3C and DIM showed a positive modulation of glucose, insulin, hemoglobin and glycated hemoglobin levels. In the same time a decreased levels of different mediators of oxidative stress were noticed, including

thiobarbituric acid reactive substances (TBARS), lipid hydroperoxides (LOOH) and conjugated dienes. Simultaneously, in this diabetic mouse model increased levels of antioxidant enzymes and small molecules (SOD, CAT, GPx, vitamin C, vitamin E, GSH) were demonstrated. Interestingly, the antioxidant action was comparable to that of metformin, a standard drug in diabetes 2 treatment [60].

## 6.5 Biological Activities of Indole-3-Carbinol in Animal Models

Animal models played a crucial role in discovering the cancer chemopreventive activity of cruciferous plants. Initially rodent chemical carcinogenesis models were used to assess anti-carcinogenic activity of minor dietary components, including I3C. Currently genetically modified animals, mentioned in the previous sections, allow to assess detailed mechanisms of their biological activity.

In the very first experiments of Wattenberg and his co-workers, benzo[a]pyrene induced model of lung and forestomach cancer in mice and dimethylbenz[a]anthracene induced breast neoplasia in rats were used. In these models I3C when given prior to carcinogen inhibited the formation of tumors [122]. This effect was linked to modulation of phase I enzymatic systems, namely cytochrome P450 dependent monooxygenases, involved in carcinogens activation [123, 124]. Later, several studies using animals carcinogenesis model of liver, colon, and tongue confirmed the anti-initiating activity of I3C. However, these studies have also provided evidence for promotional activity of I3C. For example, whereas I3C pretreatment and co-treatment with liver carcinogen aflatoxin B<sub>1</sub> (AFB<sub>1</sub>) strongly inhibited AFB<sub>1</sub> initiated hepatocarcinogenesis, posttreatment with I3C was strongly promotional [35].

The modulation of cytochromes P450 is also linked with potential protection against breast cancer. On the other hand, the same mechanism is probably responsible for uterine-induced cancer via upregulation of CYP1B1 and increased precancerous 4-hydroxyestrogen concentration [133]. The increase of carcinogenic 4-hydroxyestrogen following oral administration of I3C were documented also by the other studies [56]; reviewed in [15]. These observations led to conclusion that DIM showing higher bioavailability and reducing 4-hydroxyestrogen production should be recommended as an alternative to I3C in potential chemopreventive supplementation [15]. More recent studies have confirmed this suggestion that DIM was more effective in prevention prostate cancer in the transgenic adenocarcinoma mouse prostate (TRAMP) mice model than I3C [25].

Nevertheless, the more recent studies using “traditional” mouse models or transgenic animals documented that I3C has been responsible for a decrease of incidences of carcinogen-induced lung cancer [64, 88, 102], cervical cancer in HPV gene transgenic mice [61], and UV-induced skin cancer [29].

Moreover, it was shown that in rats bearing the 13762 mammary carcinoma, addition of I3C to the diet for 6 days prior to antitumor drug ET-743 (trabectedin) administration almost completely abolished manifestations of hepatotoxicity [41]. These observation further supports the concept that I3C or DIM protecting against specific cancer may be used in adjuvant therapy to overcome side-effects of conventional therapy.

Specific rodent models like mouse carotid artery injury were developed and used to assess I3C or DIM protection against cardiovascular diseases, obesity, or diabetes. As it was described in previous section, generally the results of these studies suggest that I3C has a potential benefit in preventing obesity and metabolic disorders, and the action of I3C *in vivo* may involve multiple mechanisms including decreased adipogenesis and inflammation, along with activated thermogenesis.

## 6.6 Biological Activities of Indole-3-Carbinol in Humans

Promising results of the most studies obtained in human cancer cell lines and in animal models prompted the clinical trials dietary intervention studies to evaluate the effect of I3C or DIM in risk group of patients or/and volunteers. A major focus of these trials has been on modulation of hormones metabolism. The urinary estrogen metabolite ratio of 2-hydroxyesterone to 16 $\alpha$ -hydroxyestrone was used in most of the trials as the surrogate endpoint biomarker.

The validity of this endpoint biomarker was confirmed in the early randomized clinical trials [14] in which 20 healthy subjects received 400 mg/day of I3C for 3 months. In most of the enrolled subjects I3C increased the 2-hydroxyestrone to estriol (a precursor of 16 $\alpha$ -hydroxyestrone) ratio in sustained manner without detectible side-effects, although some individuals were resistant to such change. In another trial women at increased risk for breast cancer were administered with different doses (range 50–400 mg/day) of I3C for 4 weeks. The results of this study suggested that the minimum effective dose schedule of 300 mg/day is optimal for breast cancer prevention, although should be confirmed by long-term breast cancer prevention trial [128].

In subsequent studies by Bradlow group [91] urine samples were collected from healthy subjects before and after oral ingestion of 6–7 mg/kg per day for 1 week (7 men) or 2 months (10 women). Analysis of 13 estrogen profiles supported the hypothesis that I3C induces estrogen 2-hydroxylation resulting in decreased concentrations of metabolites known to activate the estrogen receptor and suggested that I3C may have chemopreventive activity against breast cancer in humans. Later, phase I trial with women with a high-risk breast cancer were enrolled, subjects ingested 400 mg I3C daily for 4 weeks followed by a 4 week period of 800 mg I3C daily [105]. The maximal ratio increase of the urinary 2-hydroxyestrone to 16 $\alpha$ -hydroxyestrone was observed with the 400 mg daily dose of I3C, with no further increase found at 800 mg daily. Beside confirmation of the optimal dose of

I3C, these studies showed the induction of CYP1A2 which was mirrored by increase of 2-hydroxyestrone to 16 $\alpha$ -hydroxyestrone ratio, and GST.

Cumulative evidence on conversion of I3C to DIM in cell culture, peritoneal and oral use as well as substantial direct activity seen with DIM led to conclusion that there is no longer the case for considering I3C to be directly active, and rather DIM should be considered as a chemopreventive compound of choice [15]. A pilot study on the effect of BR-DIM on urinary hormone metabolites in postmenopausal women with a history of early-stage breast cancer showed a significant increase in levels of 2-hydroxyestrone as result of treatment with only 108 mg DIM/day for 30 days, however, nonsignificant increase (1.46–2.14) of 2-hydroxyestrone to 16 $\alpha$ -hydroxyestrone was noted [33]. In another study cohorts of 3–6 patients castrate-resistant, non-metastatic prostate cancer received escalating oral doses twice daily of BR-DIM 75 mg, then 150, 225, and 300 mg. Based on the results of this trial 225 mg BR-DIM dose twice daily was recommended for phase II trial. However, modest efficacy of DIM was demonstrated [53].

Cervical intraepithelial neoplasia (CIN) is a precancerous lesion of cervix. When patients with biopsy proven CIN grade II or III were treated orally with 200, or 400 mg/day of I3C for 12 weeks 50 % of them had complete regression based on their 12-week biopsy. Moreover, 2-hydroxyestrone to 16 $\alpha$ -hydroxyestrone ratio have changed in a dose-dependent manner [11]. The significant improvement in confirmed CIN I or II grade was also observed as a result of oral treatment with 2 mg/kg/day of DIM for 12 weeks. Moreover, at median follow-up of 6 months there was no statistically significant difference in any of the measured outcome between the DIM and placebo group [40].

Since the incidence of thyroid cancer is 4–5 times higher in women than in men, estrogens were suggested to contribute the pathogenesis of thyroid proliferative disease (TPD). In limited (7 patients) phase I clinical trial patients with TPD were administered with 300 mg of DIM per day for 14 days. DIM was detectable in thyroid tissue, and the ratio of 2-hydroxyestrone to 16 $\alpha$ -hydroxyestrone was increased. These results suggested that DIM can manifest the anti-estrogenic activity in situ to modulate TPD [104].

Although major focus of cancer prevention clinical trials of I3C or DIM has been concentrated on chemoprevention of hormone-dependent cancers, there were also clinical trials performed in order to evaluate indoles effect on pulmonary cancers. In this regard in phase I clinical trial patients with recurrent respiratory papillomatosis (RRP) were treated orally with I3C and had minimum follow-up of 8 months. Thirty-three percent of the study patients had a cessation of their papilloma growth and had not required surgery since the start of the study [107]. Subsequent long-term clinical trial performed by the same research group confirmed the preliminary observation indicating that I3C may be a treatment option for RRP [108]. The case of successful use of intralesional and intravenous cidofovir in association with I3C in 8-year-old girl with pulmonary papillomatosis was also reported [38].

As it was mentioned in the previous sections of this chapter, recently a large amount of evidence has demonstrated that epigenetic alterations, such as DNA



methylation, histone modifications, and non-coding miRNAs consistently contribute to carcinogenesis, and constituents in the diet, including dietary glucosinolate derivatives, have the potential to alter a number of these epigenetic events [46]. Different studies on cancer also have shown that miRNAs interact with genes in many different cellular pathways, displaying a differential gene expression profile between normal and tumor tissues and between tumor types [17]. Interestingly, interventions including BR-DIM in prostate cancer patients prior to radical prostatectomy showed re-expression of miR-34a, which was consistent with decreased expression of androgen receptor, prostate specific antigen (PSA), and Notch-1 in tissue specimens [70]. These results suggest that BR-DIM could be useful for the inactivation of androgen receptor, critically important during the development and progression of prostate cancer and thus its treatment.

Thus far, seven clinical studies have been registered using I3C and twelve using DIM ([www.clinicaltrials.gov](http://www.clinicaltrials.gov); accessed Dec 26, 2015). Four studies registered for I3C treatment have been completed for patients with prostate and breast cancers and one dietary intervention for healthy participants targeting unspecified adult solid tumors. One trial aiming at I3C effects on estrogen metabolism in obese volunteers had to be terminated because of slow accrual in the high BMI group. Among twelve studies registered for DIM, six have been completed for patients with prostate, breast, and cervical cancers as well as healthy volunteers. Trial aiming at new therapy of laryngeal papilloma in children was terminated because of lack of sufficient enrollment. Although the results of these trials have not been published yet, they assure the further extensive prospective studies on chemopreventive and/or chemotherapeutic potential of I3C and its condensation product.

## 6.7 Summary and Conclusions

It is well known that in populations which consume higher amounts of cruciferous vegetables lower incidence rate of cancer occurs or improved biochemical parameters, such as decreased oxidative stress are noticed [46, 117, 118, 121]. These effects are in part due to the biological activity of I3C and its condensation products, particularly DIM.

A wide range of cellular pathways are regulated by both indoles. Thus, many additional targets for indoles could be identified in the future using *in vitro* cell cultures and *in vivo* transgenic animal models and explain a unique anti-inflammatory and endocrine modulating activity of I3C. Although most of current data support the role of I3C and DIM in prevention of hormone-dependent cancers, it seems that their application in prevention of the other cancer as well as cardiovascular diseases, obesity, and diabetes reduction is also possible.

Experimental *in vitro* and *in vivo* studies and clinical trials performed so far, showed that I3C is a rather safe dietary supplement. However, since the long-term effects of I3C supplementation in humans are still not clear and due to some contradictory effects of I3C in animal models, the general use of I3C and DIM

supplements should be restricted until potential risks and benefits are better characterized. Taking into consideration higher activity of DIM, particularly in BR form, in comparison with I3C in term of potency and time required to obtain the effect, this I3C dimer might be a better alternative as chemopreventive supplement. Important aspect of possible clinical application of both indoles is their drug and radio-sensitization. Emerging new technologies allowing deeper inside in the mechanism of these glucosinolate derivatives activity should help to better explore this aspect.

## References

1. Abdelrahim M, Ewman K, Vanderlaag K et al (2006) 3,3'-diindolylmethane (DIM) and its derivatives induce apoptosis in pancreatic cancer cells through endoplasmic reticulum stress-dependent upregulation of DR5. *Carcinogenesis* 27:717–728
2. Aggarwal BB, Ichikawa H (2005) Molecular targets and anticancer potential of indole-3-carbinol and its derivatives. *Cell Cycle* 4:1201–1215
3. Ahmad A, Sarkar WA, Rahman KMW (2011) Role of nuclear factor-kappa B signaling in anticancer properties of indole compounds. *J Exp Clin Med* 3:55–62
4. Ahmad A, Biersack B, Li Y et al (2013) Targeted regulation of PI3 K/Akt/mTOR/NF- $\kappa$ B signaling by indole compounds and their derivatives: mechanistic details and biological implications for cancer therapy. *Anticancer Agents Med Chem* 13:1002–1013
5. Anderto MJ, Manson MM, Verschoyle RD et al (2004) Pharmacokinetics and tissue disposition of indole-3-carbinol and its acid condensation products after oral administration to mice. *Clin Cancer Res* 10:5233–5241
6. Anderton MJ, Manson MM, Verschoyle R et al (2004) Physiological modeling of formulated and crystalline 3,3'-diindolylmethane pharmacokinetics following oral administration in mice. *Drug Metab Dispos* 32:632–638
7. Arora A, Seth K, Kalra N, Shukla Y (2005) Modulation of P-glycoprotein-mediated multidrug resistance in K562 leukemic cells by indole-3-carbinol. *Toxicol Appl Pharmacol* 202:237–243
8. Arora A, Shukla Y (2003) Modulation of vinca-alkaloid induced P-glycoprotein expression by indole-3-carbinol. *Cancer Lett* 189:167–173
9. Arneson DW, Hurwitz A, McMahon LM et al (1999) Presence of 3,3'-diindolylmethane in human plasma after oral administration of indole-3-carbinol. *Proc Am Assoc Cancer Res* 40:429
10. Beaver LM, Yu TW, Sokolowski EI et al (2012) 3,3'-Diindolylmethane, but not indole-3-carbinol, inhibits histone deacetylase activity in prostate cancer cells. *Toxicol Appl Pharmacol* 263:345–351
11. Bell MC, Crowley-Nowick P, Bradlow HL et al (2000) Placebo-controlled trial of indole-3-carbinol in the treatment of CIN. *Gynecol Oncol* 78:123–129
12. Bhuiyan MM, Li Y, Banerjee S et al (2006) Down-regulation of androgen receptor by 3,3'-diindolylmethane contributes to inhibition of cell proliferation and induction of apoptosis in both hormone-sensitive LNCaP and insensitive C4-2B prostate cancer cells. *Cancer Res* 66:10064–10072
13. Bonnesen C, Eggleston IM, Hayes JD (2001) Dietary indoles and isothiocyanates that are generated from cruciferous vegetables can both stimulate apoptosis and confer protection against DNA damage in human colon cell lines. *Cancer Res* 61:6120–6130
14. Bradlow HL, Michnovicz JJ, Halper M et al (1994) Long-term responses of women to indole-3-carbinol or a high fiber diet. *Cancer Epidemiol Biomarkers Prev* 3:591–595

15. Bradlow HL (2008) Review. Indole-3-carbinol as a chemoprotective agent in breast and prostate cancer. *In Vivo* 22:441–445
16. Bray F, Ren JS, Masuyer E et al (2013) Global estimates of cancer prevalence for 27 sites in the adult population in 2008. *Int J Cancer* 132:1133–1145
17. Brait M, Sidransky D (2011) Cancer epigenetics: above and beyond. *Toxicol Mech Methods* 21:275–288
18. Brew CT, Aronchik I, Hsu JC et al (2006) Indole-3-carbinol activates the ATM signaling pathway independent of DNA damage to stabilize p53 and induce G1 arrest of human mammary epithelial cells. *Int J Cancer* 118:857–868
19. Chang HP, Wang ML, Hsu CY et al (2011) Suppression of inflammation-associated factors by indole-3-carbinol in mice fed high-fat diets and in isolated, co-cultured macrophages and adipocytes. *Int J Obes* 35:1530–1538
20. Chen DZ, Qi M, Auburn KJ et al (2001) Indole-3-carbinol and diindolylmethane induce apoptosis of human cervical cancer cells and in murine HPV16-transgenic preneoplastic cervical epithelium. *J Nutr* 131:3294–3302
21. Chen D, Banerjee S, Cui QC et al (2012) Activation of AMP-activated protein kinase by 3,3'-Diindolylmethane (DIM) is associated with human prostate cancer cell death in vitro and in vivo. *PLoS ONE* 7:e47186
22. Chen D, Carter TH, Auburn KJ (2004) Apoptosis in cervical cancer cells: implications for adjunct anti-estrogen therapy for cervical cancer. *Anticancer Res* 24:2649–2656
23. Chinni SR, Li Y, Upadhyay S et al (2001) Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene* 20:2927–2936
24. Chinni SR, Sarkar FH (2002) Akt inactivation is a key event in indole-3-carbinol-induced apoptosis in PC-3 cells. *Clin Cancer Res* 8:1228–1236
25. Cho HJ, Park SY, Kim EJ et al (2011) 3,3'- diindolylmethane inhibits prostate cancer development in the transgenic adenocarcinoma mouse prostate model. *Mol Carcinog* 50:100–112
26. Choi Y, Kim Y, Park S et al (2012) Indole-3-carbinol prevents diet-induced obesity through modulation of multiple genes related to adipogenesis, thermogenesis or inflammation in the visceral adipose tissue of mice. *J Nutr Biochem* 23:1732–1739
27. Choi Y, Um SJ, Park T (2013) Indole-3-carbinol directly targets SIRT1 to inhibit adipocyte differentiation. *Int J Obes (Lond)* 37:881–884
28. Christensen JG, LeBlanc GA (1996) Reversal of multidrug resistance in vivo by dietary administration of the phytochemical indole-3-carbinol. *Cancer Res* 56:574–581
29. Cope RB, Loehr C, Dashwood R et al (2006) Ultraviolet radiation-induced non-melanoma skin cancer in the Crl:SKH1:hr-BR hairless mouse: augmentation of tumor multiplicity by chlorophyllin and protection by indole-3-carbinol. *Photochem Photobiol Sci* 5(5):499–507
30. Cover MC, Hsieh SJ, Tran SH et al (1998) Indole-3-carbinol inhibits the expression of cyclin-dependent kinase-6 and induces a G1 cell cycle arrest of human breast cancer cells independent of estrogen receptor signaling. *J Biol Chem* 273:3838–3847
31. Cover CM, Hsieh SJ, Cram EJ et al (1999) Indole-3-carbinol and tamoxifen cooperate to arrest the cell cycle of MCF-7 human breast cancer cells. *Cancer Res* 59:1244–1251
32. Cram EJ, Liu BD, Bjeldanes LF et al (2001) Indole-3-carbinol inhibits CDK6 expression in human MCF-7 breast cancer cells by disrupting Sp1 transcription factor interactions with a composite element in the CDK6 gene promoter. *J Biol Chem* 276:22332–22340
33. Dalessandri KM, Firestone GL, Fitch MD et al (2004) Pilot study: effect of 3, 3'- diindolylmethane supplements on urinary hormone metabolites in postmenopausal women with a history of early-stage breast cancer. *Nutr Cancer* 50:161–167
34. Dashwood RH, Fong AT, Arbogast DN et al (1994) Anticarcinogenic activity of indole-3-carbinol acid products: ultrasensitive bioassay by trout embryo microinjection. *Cancer Res* 54:3617–3619
35. Dashwood RH (1998) Indole-3-carbinol: anticarcinogen or tumor promoter in brassica vegetables? *Chem Biol Interact* 110:1–5

36. Deng W, Zong J, Bian Z et al (2013) Indole-3-carbinol protects against pressure overload induced cardiac remodeling via activating AMPK- $\alpha$ . *Mol Nutr Food Res* 57:1680–1687
37. Denis LJ, Griffiths K (2000) Endocrine treatment in prostate cancer. *Semin Surg Oncol* 18:52–74
38. de Bilderling G, Bodart E, Lawson G et al (2005) Successful use of intralesional and intravenous Cidofovir in association with indole-3-carbinol in an 8-year-old girl with pulmonary papillomatosis. *J Med Virol* 75:332–335
39. De Kruif CA, Marsman JW, Venekamp JC et al (1991) Structure elucidation of acid reaction products of indole-3-carbinol: detection in vivo and enzyme induction in vitro. *Chem Biol Interact* 80:303–315
40. Del Priore G, Gudipudi DK, Montemarano N et al (2010) Oral diindolylmethane (DIM): pilot evaluation of a nonsurgical treatment for cervical dysplasia. *Gynecol Oncol* 116:464–467
41. Donald S, Verschoyle RD, Greaves P et al (2004) Dietary agent indole-3-carbinol protects female rats against the hepatotoxicity of the antitumor drug ET-743 (trabectedin) without compromising efficacy in a rat mammary carcinoma. *Int J Cancer* 111:961–967
42. Dunn SE, LeBlanc GA (1994) Hypocholesterolemic properties of plant indoles. Inhibition of acyl-CoA:cholesterol acyltransferase activity and reduction of serum LDL/VLDL cholesterol levels by glucobrassicin derivatives. *Biochem Pharmacol* 47:359–364
43. Dzau VJ, Braun-Dullaeus RC, Sedding DG (2002) Vascular proliferation and atherosclerosis: new perspectives and therapeutic strategies. *Nat Med* 8:1249–1256
44. Fan S, Meng Q, Auburn K et al (2006) BRCA1 and BRCA2 as molecular targets for phytochemicals indole-3-carbinol and genistein in breast and prostate cancer cells. *Br J Cancer* 94:407–426
45. Fares F (2014) The anti-carcinogenic effect of indole-3-carbinol and 3,3'-diindolylmethane and mechanism of action. *Med Chem*. doi:10.4172/2161-0444.S1-002
46. Fuentes F, Paredes-Gonzalez X, Kong AT (2015) Dietary glucosinolates sulforaphane, phenethyl isothiocyanate, indole-3-carbinol/3,3'-diindolylmethane: anti-oxidative stress/inflammation, nrf2, epigenetics/epigenomics and in vivo cancer chemopreventive efficacy. *Curr Pharmacol Rep* 1:179–196
47. Fujioka N, Ainslie-Waldman CE, Upadhyaya P et al (2014) Urinary 3,3'-diindolylmethane: a biomarker of glucobrassicin exposure and indole-3-carbinol uptake in humans. *Cancer Epidemiol Biomark Prev* 23:282–287
48. Garcia HH, Brar GA, Nguyen DH et al (2005) Indole-3-Carbinol (I3C) inhibits cyclin-dependent kinase-2 function in human breast cancer cells by regulating the size distribution, associated cyclin E forms, and subcellular localization of the CDK2 protein complex. *J Biol Chem* 280:8756–8764
49. Garikapaty VP, Ashok BT, Chen YG et al (2005) Anti-carcinogenic and anti-metastatic properties of indole-3-carbinol in prostate cancer. *Oncol Rep* 13:89–93
50. Ge X, Fares FA, Yannai S (1999) Induction of apoptosis in MCF-7 cells by indole-3-carbinol is independent of p53 and bax. *Anticancer Res* 19:3199–3203
51. Guan H, Chen C, Zhu L et al (2013) Indole-3-carbinol blocks platelet-derived growth factor-stimulated vascular smooth muscle cell function and reduces neointima formation in vivo. *J Nutr Biochem* 24:62–69
52. Hayes JD, Dinkova-Kostova AT, McMahon M (2009) Cross-talk between transcription factors AhR and Nrf2: lessons for cancer chemoprevention from dioxin. *Toxicol Sci* 111:199–201
53. Heath EI, Heilbrun LK, Li J (2010) Phase I dose-escalation study of oral BR-DIM (BioResponse 3,3'-Diindolylmethane) in castrate-resistant, non-metastatic prostate cancer. *Am J Transl Res* 2:402–411
54. Heinlein CA, Chang C (2004) Androgen receptor in prostate cancer. *Endocr Rev* 25:276–308
55. Hong C, Kim HA, Firestone GL et al (2002) 3,3'-Diindolylmethane (DIM) induces a G(1) cell cycle arrest in human breast cancer cells that is accompanied by Sp1-mediated activation of p21(WAF1/CIP1) expression. *Carcinogenesis* 23:1297–1305

56. Horn TL, Reichert MA, Bliss RL (2002) Modulations of P450 mRNA in liver and mammary gland and P450 activities and metabolism of estrogen in liver by treatment of rats with indole-3-carbinol. *Biochem Pharmacol* 64:393–404
57. Hwang JW, Jung JW, Lee YS et al (2008) Indole-3-carbinol prevents H(2)O(2)-induced inhibition of gap junctional intercellular communication by inactivation of PKB/Akt. *J Vet Med Sci* 70:1057–1063
58. International Agency for Research on Cancer (1999) Monographs on the evolution of carcinogenic risks to humans: hormonal contraception and postmenopausal hormone therapy, vol 72. IARC, Lyon, France
59. Izzotti A, Calin GA, Steele VE et al (2010) Chemoprevention of cigarette smoke-induced alterations of microRNA expression in rat lungs. *Cancer Prev Res* 3:62–72
60. Jayakumar P, Pugalendi KV, Sankaran M (2014) Attenuation of hyperglycemia-mediated oxidative stress by indole-3-carbinol and its metabolite 3, 3'- diindolylmethane in C57BL/6 J mice. *J Physiol Biochem* 70:525–534
61. Jin L, Qi M, Chen DZ et al (1999) Indole-3-carbinol prevents cervical cancer in human papilloma virus type 16 (HPV16) transgenic mice. *Cancer Res* 59:3991–3997
62. Jin Y (2011) 3,3'-Diindolylmethane inhibits breast cancer cell growth via miR-21-mediated Cdc25A degradation. *Mol Cell Biochem* 358:345–354
63. Kassie F, Anderson LB, Scherber R et al (2007) Indole-3-carbinol inhibits 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone plus benzo(a)pyrene-induced lung tumorigenesis in A/J mice and modulates carcinogen-induced alterations in protein levels. *Cancer Res* 67:6502–6511
64. Kassie F, Kalscheuer S, Matisse I et al (2010) Inhibition of vinyl carbamate-induced pulmonary adenocarcinoma by indole-3-carbinol and myo-inositol in A/J mice. *Carcinogenesis* 31:239–245
65. Kassie F, Melkamu T, Endalew A et al (2010) Inhibition of lungcarcinogenesis and critical cancer-related signaling pathways by N-acetyl-S-(N-2-phenethylthiocarbamoyl)-l-cysteine, indole-3-carbinol and myo-inositol, alone and in combination. *Carcinogenesis* 31:1634–1641
66. Kim DJ, Han BS, Ahn B et al (1997) Enhancement by indole-3-carbinol of liver and thyroid gland neoplastic development in a rat medium-term multiorgan carcinogenesis model. *Carcinogenesis* 18:377–381
67. Kim EJ, Park Sy, Shin et al (2007) Activation of caspase-8 contributes to 3,3'-Diindolylmethane-induced apoptosis in colon cancer cells. *J Nutr* 137:31–36
68. Kojima T, Tanaka T, Mori H (1994) Chemoprevention of spontaneous endometrial cancer in female Donryu rats by dietary indole-3-carbinol. *Cancer Res* 54:1446–1449
69. Kong D, Heath E, Chen W et al (2012) Loss of let-7 up-regulates EZH2 in prostate cancer consistent with the acquisition of cancer stem cell signatures that are attenuated by BR-DIM. *PLoS ONE* 7:e33729
70. Kong D, Heath E, Chen W et al (2012) Epigenetic silencing of miR-34a in human prostate cancer cells and tumor tissue specimens can be reversed by BR-DIM treatment. *Am J Transl Res* 4:14–23
71. Kumi-Diaka J, Merchant K, Haces A et al (2010) Genistein-selenium combination induces growth arrest in prostate cancer cells. *J Med Food* 13:842–850
72. Kumar MM, Davuluri S, Poojar S et al (2015) Role of estrogen receptor alpha in human cervical cancer-associated fibroblasts: a transcriptomic study. *Tumour Biol Oct 24* [Epub ahead of print]
73. Lawrence T (2009) The nuclear factor NF- $\kappa$ B pathway in Inflammation. *Cold Spring Harb Perspect Biol* 1:a001651. doi:10.1101/cshperspect.a001651
74. Le HT, Schaldach CM, Firestone GL et al (2003) Plant-derived 3,3'-Diindolylmethane is a strong androgen antagonist in human prostate cancer cells. *J Biol Chem* 278:21136–21145
75. Leong H, Riby JE, Firestone GL et al (2004) Potent ligand-independent estrogen receptor activation by 3,3'-diindolylmethane is mediated by cross talk between the protein kinase A and mitogen-activated protein kinase signaling pathways. *Mol Endocrinol* 18:291–302

76. Li Y, Li X, Sarkar FH (2003) Gene expression profiles of I3C- and DIM-treated PC3 human prostate cancer cells determined by cDNA microarray analysis. *J Nutr* 133:1011–1019
77. Li Y, Wang Z, Kong D et al (2007) Regulation of FOXO3a/beta-catenin/GSK-3beta signaling by 3,3'-diindolylmethane contributes to inhibition of cell proliferation and induction of apoptosis in prostate cancer cells. *J Biol Chem* 282:21542–21550
78. Li Y, VandenBoom II TG, Wang Z et al (2010) miRNA146a suppresses invasion of pancreatic cancer cells. *Cancer Res* 70:1486–1495
79. Lian JP, Word B, Taylor S et al (2004) Modulation of the constitutive activated STAT3 transcription factor in pancreatic cancer prevention: effects of indole-3-carbinol (I3C) and genistein. *Anticancer Res* 24:133–137
80. Licznarska BE, Szaefer H, Murias M et al (2013) Modulation of CYP19 expression by cabbage juices and their active components: indole-3-carbinol and 3,3'-diindolylmethane in human breast epithelial cell lines. *Eur J Nutr* 52:1483–1492
81. Lo R, Matthews J (2013) The aryl hydrocarbon receptor and estrogen receptor-alpha differentially modulate nuclear factor erythroid-2-related factor2 transactivation in MCF-7 breast cancer cells. *Toxicol Appl Pharmacol* 270:139–148
82. Lu Q, Nakamura J, Savinov A et al (1996) Expression of aromatase protein and messenger ribonucleic acid in tumor epithelial cells and evidence of functional significance of locally produced estrogen in human breast cancer. *Endocrinology* 137:3061–3068
83. Luo J, Manning BD, Cantley LC (2003) Targeting the PI3 K-Akt pathway in human cancer: rationale and promise. *Cancer Cell* 4:257–262
84. Lynn A, Collins A, Fuller Z et al (2006) Cruciferous vegetables and colorectal cancer. *Proc Nutr Soc* 65:135–144
85. Maiyoh GK, Kuh JE, Casaschi A et al (2007) Cruciferous indole-3-carbinol inhibits apolipoprotein B secretion in HepG2 cells. *J Nutr* 137:2185–2189
86. Marconett CN, Singhal AK, Sundar SN et al (2012) Indole-3-carbinol disrupts estrogen receptor-alpha dependent expression of insulin-like growth factor-1 receptor and insulin receptor substrate-1 and proliferation of human breast cancer cells. *Mol Cell Endocrinol* 363:74–84
87. McGuire KP, Ngoubilly N, Neavyn M et al (2006) 3,3'-diindolylmethane and paclitaxel act synergistically to promote apoptosis in HER2/Neu human breast cancer cells. *J Surg Res* 132:208–213
88. Melkamu T, Zhang X, Tan J et al (2010) Alteration of microRNA expression in vinyl carbamate-induced mouse lung tumors and modulation by the chemopreventive agent indole-3-carbinol. *Carcinogenesis* 31:252–258
89. Meng Q, Qi M, Chen DZ et al (2000) Suppression of breast cancer invasion and migration by indole-3-carbinol: associated with up-regulation of BRCA1 and E-cadherin/catenin complexes. *J Mol Med* 78:155–165
90. Mesnil M, Crespin S, Avanzo JL et al (2005) Defective gap junctional intercellular communication in the carcinogenic process. *Biochim Biophys Acta* 1719:125–145
91. Michnovicz JJ, Adlercreutz H, Bradlow HL (1997) Changes in levels of urinary estrogen metabolites after oral indole-3-carbinol treatment in humans. *J Natl Cancer Inst* 89:718–723
92. Mulvey L, Chandrasekaran A, Liu K et al (2007) Interplay of genes regulated by estrogen and diindolylmethane in breast cancer cell lines. *Mol Med* 13:69–78
93. Nakamura Y, Yogosawa S, Izutani Y et al (2009) A combination of indole-3-carbinol and genistein synergistically induces apoptosis in human colon cancer HT-29 cells by inhibiting Akt phosphorylation and progression of autophagy. *Mol Cancer* 8:100. doi:10.1186/1476-4598-8-100
94. Nachshon-Kedmi M, Yannai S, Haj A et al (2003) Indole-3-carbinol and 3,3'-diindolylmethane induce apoptosis in human prostate cancer cells. *Food Chem Toxicol* 41:745–752
95. Oganessian A, Hendricks JD, Williams DE (1997) Long term dietary indole-3-carbinol inhibits diethylnitrosamine-initiated hepatocarcinogenesis in the infant mouse model. *Cancer Lett* 118:87–94

96. Ohtake F, Fujii-Kuriyama Y, Kawajiri K et al (2011) Cross-talk of dioxin and estrogen receptor signals through the ubiquitin system. *J Steroid Biochem Mol Biol* 127:102–107
97. Pagliaro B, Santolamazza C, Simonelli F et al (2015) Phytochemical compounds and protection from cardiovascular diseases: a state of the art. *BioMed Res Int*. doi:[10.1155/2015/918069](https://doi.org/10.1155/2015/918069)
98. Paik WH, Kim HR, Park JK et al (2013) Chemosensitivity induced by down-regulation of MicroRNA-21 in gemcitabine-resistant pancreatic cancer cells by indole-3-carbinol. *Anticancer Res* 33:1473–1482
99. Park MK, Rhee YH, Lee HJ et al (2008) Antiplatelet and antithrombotic activity of indole-3-carbinol in vitro and in vivo. *Phytother Res* 22:58–64
100. Pearson G, Robinson F, Beers Gibson T et al (2001) Mitogen-activated protein (MAP) kinase pathways: regulation and physiological functions. *Endocr Rev* 22:153–183
101. Penning TM, Burczynski ME, Jez JM et al (2000) Human 3 $\alpha$ -hydroxysteroid dehydrogenase isoforms (AKR1C1-AKR1C4) of the aldo-keto reductase superfamily: functional plasticity and tissue distribution reveals roles in the inactivation and formation of male and female sex hormones. *Biochem J* 351:67–77
102. Qian X, Melkamu T, Upadhyaya P et al (2011) Indole-3-carbinol inhibited tobacco smokecarcinogen-induced lung adenocarcinoma in A/J mice when administered during the post-initiation or progression phase of lung tumorigenesis. *Cancer Lett* 311:57–65
103. Rahman KM, Aranha O, Sarkar FH (2003) Indole-3-carbinol (I3C) induces apoptosis in tumorigenic but not in nontumorigenic breast epithelial cells. *Nutr Cancer* 45:101–112
104. Rajoria S, Suriano R, Parmar PS et al (2011) 3,3'-diindolylmethane modulates estrogen metabolism in patients with thyroid proliferative disease: a pilot study. *Thyroid* 21:299–304
105. Reed GA, Peterson KS, Smith HJ et al (2005) A phase I study of indole-3-carbinol in women: tolerability and effects. *Cancer Epidemiol Biomark Prev* 14:1953–1960
106. Rice JC, Ozcelik H, Maxeiner P et al (2000) Methylation of the BRCA1 promoter is associated with decreased BRCA1 mRNA levels in clinical breast cancer specimens. *Carcinogenesis* 21:1761–1765
107. Rosen CA, Woodson GE, Thompson JW et al (1998) Preliminary results of the use of indole-3-carbinol for recurrent respiratory papillomatosis. *Otolaryngol Head Neck Surg* 118:810–815
108. Rosen CA, Bryson PC (2004) Indole-3-carbinol for recurrent respiratory papillomatosis: long-term results. *J Voice* 18:248–253
109. Sarkar FH, Li Y, Wang Z et al (2009) Cellular signaling perturbation by natural products. *Cell Signal* 21:1541–1547
110. Sarkar FH, Li Y (1997) Indole-3-carbinol and prostate cancer. *J Nutr* 134:3493S–3498S
111. Sarkar S, Dubaybo H, Ali S et al (2013) Down-regulation of miR-221 inhibits proliferation of pancreatic cancer cells through up-regulation of PTEN, p27(kip1), p57(kip2), and PUMA. *Am. J Cancer Res* 3:465–477
112. Shaw RJ, Cantley LC (2006) Ras, PI(3)K and mTOR signalling controls tumour cell growth. *Nature* 441:424–430
113. Singhal R, Shankar K, Badger TM et al (2008) Estrogenic status modulates aryl hydrocarbon receptor-mediated hepatic gene expression and carcinogenicity. *Carcinogenesis* 29:227–236
114. Szaefer H, Krajka-Kuźniak V, Licznarska B (2015) Cabbage juices and indoles modulate the expression profile of AhR, ER $\alpha$ , and Nrf2 in human breast cell lines. *Nutr Cancer* 67:1342–1345
115. Śmiechowska A, Bartoszek A, Namieśnik J (2008) Cancer chemopreventive agents: Glucosinolates and their decomposition products in white cabbage (*Brassica oleracea* var. *Capitata*). *Postepy Hig Med Dosw* (online) 62:125–140
116. Tadi K, Chang Y, Ashok BT et al (2005) 3,3'-Diindolylmethane, a cruciferous vegetable derived synthetic antiproliferative compound in thyroid disease. *Biochem Biophys Res Commun* 337:1019–1025
117. Terry P, Wolk A, Persson I et al (2001) Brassica vegetables and breast cancer risk. *JAMA* 285:2975–2977

118. van Poppel G, Verhoeven DT, Verhagen H et al (1999) Brassica vegetables and cancer prevention. *Epidemiology and mechanisms*. *Adv Exp Med Biol* 472:159–168
119. Vahid F, Zand H, Nosrat-Mirshekarlou E et al (2015) The role dietary of bioactive compounds on the regulation of histone acetylases and deacetylases: a review. *Gene* 562:8–15
120. Vang O (2006) Chemopreventive potential of compounds in Cruciferous vegetables. In: Baer-Dubowska W, Bartoszek A, Malejka-Giganti D (eds) *Carcinogenic and anticarcinogenic food components*. CRC Taylor & Francis, Boca Raton, pp 303–328
121. Verhagen H, Poulsen HE, Loft S et al (1995) Reduction of oxidative DNA-damage in humans by brussels sprouts. *Carcinogenesis* 16:969–970
122. Wattenberg LW, Loub WD (1978) Inhibition of polycyclic aromatic hydrocarbon-induced neoplasia by naturally occurring indoles. *Cancer Res* 38:1410–1413
123. Wattenberg LW, Loub WD, Lam LK, Speier JL (1976) Dietary constituents altering the responses to chemical carcinogens. *Fed Proc* 35:1327–1331
124. Wattenberg LW, Hanley AB, Barany G et al (1985) Inhibition of carcinogenesis by some minor dietary constituents. *Princess Takamatsu Symp* 16:193–203
125. WHO Report Part II 2015
126. Wilson CA, Ramos L, Villaseñor MR et al (1999) Localization of human BRCA1 and its loss in high-grade, non-inherited breast carcinomas. *Nat Genet* 21:236–240
127. Witter DC, Le Bas J (2008) Cancer as a chronic disease. *Oncology* 53:1–3
128. Wong GY, Bradlow L, Sepkovic D et al (1997) Dose-ranging study of indole-3-carbinol for breast cancer prevention. *J Cell Biochem Suppl* 28–29:111–116
129. Wong CP, Hsu A, Buchanan A et al (2014) Effects of sulforaphane and 3,3'-diindolylmethane on genome-wide promoter methylation in normal prostate epithelial cells and prostate cancer cells. *PLoS ONE* 9:e86787. doi:10.1371/journal.pone.0086787
130. Wu TY, Khor TO, Su ZY et al (2013) Epigenetic modifications of Nrf2 by 3,3'-diindolylmethane in vitro in TRAMP C1 cell line and in vivo TRAMP prostate tumors. *AAPS J* 15:864–874
131. Xu M, Orner GA, Bailey GS et al (2001) Post-initiation effects of chlorophyllin and indole-3-carbinol in rats given 1,2-dimethylhydrazine or 2-amino-3-methylimidazo[4,5-f]quinoline. *Carcinogenesis* 22:309–314
132. Xu H, Barnes GT, Yang Q et al (2003) Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 112:1821–1830
133. Yoshida M, Katashima S, Ando J et al (2004) Dietary indole-3-carbinol promotes endometrial adenocarcinoma development in rats initiated with N-ethyl-N'-nitro-N-nitrosoguanidine, with induction of cytochrome P450s in the liver and consequent modulation of estrogen metabolism. *Carcinogenesis* 25:2257–2264
134. Zhang J, Hsu BAJC, Kinseth BAMA et al (2003) Indole-3-carbinol induces a G1 cell cycle arrest and inhibits prostate-specific antigen production in human LNCaP prostate carcinoma cells. *Cancer* 98:2511–2520
135. Zhu J, Li Y, Guan C et al (2012) Anti-proliferative and pro-apoptotic effects of 3, 3'-diindolylmethane in human cervical cancer cells. *Oncol Rep* 28:1063–1068